

The regeneration of damaged connective tissue: wishful thinking or reality?

U. Schneider¹, W.D. Murrell², P. Hollands³

¹iRegMed, Tegernsee, Germany

²Abu Dhabi Knee and Sports Medicine, Healthpoint Hospital, Zayed Sports City, Abu Dhabi, United Arab Emirates

³Freelance Consultant Clinical Scientist, Cambridge, UK

Corresponding Author: Peter Hollands, Ph.D (Cantab), FRSB; e-mail: peterh63@hotmail.com

Keywords: Adipose tissue, Bone marrow, Connective tissue, Cytokines, GOLDIC®, Growth factors, Mesenchymal stem cells, Platelet-rich plasma, Tissue regeneration, Tissue repair.

ABSTRACT

This review examines the current status of various cellular and non-cellular regeneration technologies used for the repair and regeneration of damaged connective tissue. The article explores the clinical use of bone marrow-derived mesenchymal stem cells, adipose tissue-derived mesenchymal stem cells, growth factors, cytokines, platelet-rich plasma and GOLDIC® method. To compare the regenerative capacity of these technologies, a systematic analysis of the regeneration quality is necessary, and a high-resolution magnetic resonance imaging (MRI) using a quality scoring system is needed. It is likely that in future clinical practice a combination of such technologies will offer the optimal treatment to patients with different connective tissue disorders, which must always be our ultimate goal.

INTRODUCTION

Connective tissues such as ligaments, tendons, intervertebral discs, and articular cartilage have a limited capacity to heal following structural damage¹. Nevertheless, bone can heal when injured thanks to the high degree of vascularization and the appropriate cellular environment to promote tissue repair². It is known that urodele amphibians

such as the newt can regenerate their tails, limbs, lens, retina, jaw, and even a large portion of the heart^{3,4}, but the capacity for regeneration of whole tissues and organs has been lost in mammals⁵. The inadequacy of true connective tissue regeneration in mammals has been attributed to the absence of blastema formation (a reverse developmental process occurring partly via cell de-differentiation in tissues local to the amputation plane and partly via a contribution of muscle stem cells) and to the rapid fibroproliferative response after wounding⁶.

The physiological healing process of the connective tissue can be broadly separated into the processes of regeneration and repair⁷. Regeneration results in the complete restitution of lost or damaged tissue, whereas repair may restore some original structures but involves collagen deposition and scar formation⁸. Chronic inflammation stimulates scar formation through local production of growth factors and cytokines that promote fibroblast proliferation and collagen synthesis⁹.

Tissue repair and regeneration depend not only on the activity of humoral factors, but also on interactions between cells and the components of the extracellular matrix (ECM)¹⁰. The ECM regulates the growth¹¹, proliferation¹², migration¹³, and differentiation¹⁴ of the cells residing within it. It is proposed that the ECM constantly undergoes remodelling in both physiological and pathological processes. The synthesis and degradation of ECM is associated with morphogenesis¹⁵, wound healing¹⁶, chronic fibrosis¹⁷, regeneration¹⁸, and metastatic processes¹⁹. The ECM components can regulate cell proliferation by signaling through cellular receptors belonging to the inte-



grin family²⁰. The type of ECM proteins can affect the degree of cell differentiation²¹, and the maintenance of normal tissue structure requires a basement membrane or stromal scaffold²². The integrity of basement membrane and parenchymal cells is critical for the organized regeneration of tissues²³. It is worth noting that tissue injury results in restitution of the normal structure only if the ECM is not damaged, although labile and stable cells are capable of regeneration. Disruption of the ECM ultimately leads to collagen deposition and scar formation²⁴.

The regenerative capacity of any tissue depends on fibroblast growth factors²⁵ and cell signaling mechanisms²⁶. Therefore, it is not surprising that regenerative therapeutic approaches are focused on the use of cells (including stem cells) and growth factors. Of note, the use of growth factors, platelet-rich plasma (PRP), autologous differentiated cells and mesenchymal stem cells (MSCs) has shown the most promise for the treatment of musculoskeletal diseases²⁷⁻³⁰. The efficacy of these treatments is based on their potential to regenerate tissues which cannot be regenerated under physiological conditions. However, the proof of concept of the efficacy of such new therapies has not been fully achieved yet³¹. Indeed, true tissue regeneration has to be proven by histology or high-resolution magnetic resonance imaging (MRI). Some technologies have demonstrated promise, but none of them has proven to induce true connective tissue regeneration, which consists of complete restitution of damaged tissue on a histological level³².

BONE MARROW-DERIVED MESENCHYMAL STEM CELLS (BM-MSCs)

Stem cells are characterized by self-renewal properties and by their capacity to differentiate into different cell lineages³³. MSCs are multipotent cells, which means that they have potentially important therapeutic applications since they can generate chondrocytes, osteoblasts, adipocytes, myoblasts, and endothelial cell precursors depending on the tissue to which they migrate³⁴. MSCs migrate to injured tissues and generate stromal cells or other cell lineages, but they do not seem to participate in normal tissue homeostasis. More recently, some authors have proposed that MSCs play an important role in normal tissue homeostasis³⁵, which may reflect the true role of MSCs in tissue homeostasis. At the

time of writing, MSCs have been isolated from bone marrow³⁶, periosteum³⁷, trabecular bone³⁸, adipose tissue³⁹, synovium⁴⁰, skeletal muscle⁴¹, deciduous and adult teeth⁴², umbilical cord blood⁴³, umbilical cord tissue⁴⁴⁻⁴⁶, placenta⁴⁷, and several other sources such as menstrual blood⁴⁸ and milk⁴⁹, which are in the early stages of research and development.

BM-MSCs can differentiate into cells belonging to the connective tissue lineage, including bone⁵⁰, fat⁵¹, cartilage⁵², intervertebral disc cells⁵³, ligaments⁵⁴, and cardiomyocytes⁵⁵. BM-MSCs generate rapidly dividing cells known as transit-amplifying cells (TACs) which lose their capacity of self-renewal and give rise to cells with restricted developmental potential known as progenitor cells⁵⁶. BM-MSCs can be isolated and expanded *in vitro* ideally using an automated bioreactor to optimize the quality and safety of the expanded cell product⁵⁷.

BM-MSCs clearly have a great potential in the treatment of damaged connective tissue, particularly in osteoarthritis⁵⁸. However, there are some problems that still need to be resolved. These problems include clinical challenges⁵⁹ and a careful consideration of regulatory issues arising from the use of expanded human stem cells for clinical applications⁶⁰ (Table 1).

ADIPOSE-DERIVED MESENCHYMAL STEM CELLS (AD-MSCs)

In general, adipose-derived mesenchymal stem cells (AD-MSCs) are characterized by a reduced expression of bone morphogenetic protein (BMP)-2, BMP-4 and BMP-6, and by the lack of expression of transforming growth factor (TGF)- β type 1 receptor when compared to BM-MSCs⁶¹. Therefore, supplementation of these factors is needed if osteogenic or chondrogenic differentiation is desired from AD-MSCs. Currently, adipose-derived cells and tissues can be prepared with increasing regenerative therapeutic potency through different strategies, including:

1. Fat grafting, which is usually associated with cosmetic and plastic/reconstructive surgery applications⁶².
2. Micronized or emulsified fat, which is commonly used in plastic and reconstructive surgery⁶³.
3. Mechanically and enzymatically processed stromal vascular fraction (SVF), which contains free AD-MSCs⁶⁴.

Table 1. Description of the different regenerative techniques for connective tissue repair and regeneration based on their regenerative capacity, regulation, side effects, risks, costs, and availability.

	BM-MSCs	AD-MSCs	Adipose SVF	Adipose Tissue	Cytokines and growth factors	PRP	GOLDIC®
Regenerative capacity	+	+	+	+	+/-	+/-	++
Regulatory requirements	ATMP	ATMP	ATMP	Medical Product	Pharmaceutical	Medical Product	Medical Product
Collection procedure	Bone marrow aspiration	Adipose tissue harvesting	Adipose tissue harvesting	Adipose tissue harvesting	None	Venepuncture	Venepuncture
Preparation	<i>In vitro</i> culture and differentiation of cells with various protocols	<i>In vitro</i> culture and differentiation of cells with various protocols	Single step aspiration and preparation (mechanical or enzymatic processing)	Single step aspiration and preparation, washing or emulsification without enzymes	Ready to use	Centrifugation of whole blood	<i>In vitro</i> culture of whole blood with defined gold particles for 24 hours
Risks	Moderate	Moderate	Moderate	Minimal	Minimal	None	None
Costs	High	High	Moderate	Moderate	Moderate	Low	Low
Availability	Veterinary market; few in human market	Veterinary market; few in human market	Worldwide (although regulatory constraints exist)	Worldwide	Worldwide	Worldwide	Veterinary market worldwide; human market in Europe

Abbreviations: AD-MSCs: adipose-derived mesenchymal stem cells; ATMP: advanced therapy medicinal products; BM-MSCs: bone marrow-derived mesenchymal stem cells; GOLDIC®: gold-induced cytokines; PRP: platelet-rich plasma; SVF: stromal vascular fraction.

- Mechanically digested SVF mixed with the ECM concentrate, which is also referred to as “stromal vascular matrix” (SVM) and combines both free AD-MSCs and associated ECM⁶⁵.
- In vitro* expanded AD-MSCs containing much higher numbers of AD-MSCs⁶⁶.

Fat graft has minimal to no potency as the MSCs are not liberated from their perivascular niche and are difficult to activate. Micronized or emulsified fat has improved biochemical activity as stem cells are found in smaller micro-niches. Mechanical washing and removal of fibrous tissue enable cells to survive longer in their implanted microenvironment. SVF and SVM require mechanical or enzymatic digestion of whole adipose tissue and separation from other cell types by centrifugation. In most countries, enzymatic adipose tissue processing results in an “advanced therapy medicinal products” (ATMP), whereas mechanical processing does not result in an ATMP making it relatively easier to use. The use of expanded AD-MSCs is considered a higher regulatory risk because of the increased manipulation of these cells,

which are therefore more strictly regulated. In general, AD-MSCs exhibit immunomodulatory⁶⁷ and trophic properties⁶⁸, and originate from local pericytes liberated from the broken blood vessels during processing. *In situ* activated AD-MSCs secrete a range of bioactive agents that locally inhibit the overactive immune system, resulting in an important line of defence against the development of autoimmune responses due to the antigen exposure following tissue injury. On the other hand, the trophic effects of AD-MSCs help to establish an optimal regenerative microenvironment at the site of injury by: i) inhibiting ischemia-related apoptosis⁶⁹, ii) downregulating scar formation⁷⁰, iii) stimulating angiogenesis via secretion of vascular endothelial growth factor (VEGF)⁷¹, iv) promoting capillary stabilization through AD-MSC-derived pericytes⁷², v) secreting tissue progenitor-specific mitogens that enhance tissue regeneration⁷³.

AD-MSCs are considerably promising for the regeneration of damaged connective tissue. However, as with BM-MSCs, there are technical, regulatory and translational issues that need to be

resolved before AD-MSCs can be brought into routine clinical use. Limitations and regulatory issues of AD-MSCs are shown in Table 1.

ROLE OF GROWTH FACTORS IN THE REGENERATION OF DAMAGED CONNECTIVE TISSUE

Growth factors have many roles in normal cellular homeostasis, including promotion of cell survival⁷⁴, induction of cell proliferation⁷⁵, and stimulation of cell contractility⁷⁶, cell locomotion⁷⁷, cell differentiation⁷⁸, and angiogenesis⁷⁹. Growth factors act as ligands by binding to specific cell surface receptors, which in turn deliver signals to the target cells. These signalling pathways stimulate gene transcription⁸⁰ that may be silent in resting cells and may involve genes that control the entry into the cell cycle⁸¹.

PLATELET-DERIVED GROWTH FACTOR (PDGF)

Platelet-derived growth factor (PDGF) constitutes a family of several closely related polypeptides, consisting of chains linked by disulphide bridges and resulting in five dimeric isoforms⁸². PDGF is stored in platelet granules and is released upon platelet activation⁸³. PDGF is also produced by a variety of cells other than platelets, and it has been shown to play an important role in bone regeneration⁸⁴.

VASCULAR ENDOTHELIAL GROWTH FACTOR (VEGF)

Vascular endothelial growth factor (VEGF) induces blood vessel formation in early development through a process known as vasculogenesis⁸⁵, and it has a central role in the growth of new blood vessels (angiogenesis) in adults⁸⁶. Specifically, vasculogenesis is defined as the differentiation of precursor cells (angioblasts) into endothelial cells and the *de novo* formation of a primitive vascular network, whereas angiogenesis is defined as the growth of new capillaries from pre-existing blood vessels⁸⁷.

VEGF promotes angiogenesis in chronic inflammation⁸⁸, wound healing⁸⁹, and tumorigenesis⁹⁰. VEGF acts via three tyrosine kinase receptors (Vascular endothelial growth factor receptors, a.k.a. VEGFRs): VEGFR-1, VEGFR-2 and VEGFR-3. VEGFR-2 is expressed in endothelial cells and many other cell types. These are the main receptors associated with the vasculogenic and angiogenic effects of VEGF⁹¹. The role of VEGFR-1 is less well understood, although it is thought to facilitate the mobilization of endothelial stem cells

and to have a role in inflammation⁹². VEGF-C and VEGF-D bind to VEGFR-3 and act on lymphatic endothelial cells to induce lymphangiogenesis⁹³. The pivotal role of VEGF in vasculogenesis, angiogenesis and lymphangiogenesis indicates that VEGF is an important component of the regenerative mechanisms for damaged connective tissue.

FIBROBLAST GROWTH FACTOR (FGF)

Fibroblast growth factor (FGF) exists in more than 20 isoforms⁹⁴. Acidic FGF (aFGF, or FGF-1)⁹⁵ and basic FGF (bFGF, or FGF-2)⁹⁶ are the best characterized FGF isoforms in terms of structure and function. Most FGF molecules transduce signals via four tyrosine kinase receptors: fibroblast growth factor receptor (FGFR)-1, FGFR-2, FGFR-3 and FGFR-4^{97,98}. FGF-1 is capable of binding to all these receptors⁹⁸. FGF-7 is also known as keratinocyte growth factor (KGF)⁹⁹. FGF signaling contributes to wound healing¹⁰⁰, angiogenesis¹⁰¹, hematopoiesis¹⁰², skeletal development¹⁰³, and many other biological processes. The wide range of biological activities exerted by FGFs suggests that these growth factors play an important role in the regeneration and repair of damaged connective tissue.

TRANSFORMING GROWTH FACTOR- β (TGF- β) AND RELATED GROWTH FACTORS

There are approximately 30 different types of transforming growth factors (TGFs) which include three TGF- β isoforms, namely: TGF- β 1, TGF- β 2 and TGF- β 3^{104,105}. TGF- β is a homodimeric protein produced by many different cell types such as platelets¹⁰⁶, lymphocytes¹⁰⁷, macrophages¹⁰⁸, and endothelial cells¹⁰⁹. TGF- β has multiple and often opposing effects depending on the tissue and the type of injury¹¹⁰; for instance, TGF- β has growth inhibition properties in most epithelial cells¹¹¹. In this regard, loss of TGF- β receptors may occur during tumorigenesis, providing a proliferative advantage to cancer cells¹¹². Overall, TGF- β seems to participate in most cellular processes and is therefore an excellent candidate molecule potentially involved in the repair of damaged connective tissue.

CYTOKINES

Cytokines have important functions as mediators of inflammation and immune responses¹¹³. Cytokines contribute to the homeostasis of bone and

connective tissue¹¹⁴ and play an important role in the regeneration of bone and connective tissue¹¹⁵. It is highly likely that cytokines play a critical role in the regeneration of damaged connective tissue and it is therefore important to consider their use in parallel with both cell- and non-cell-based therapies¹¹⁶. The potential clinical applications and limitations of the use of growth factors and cytokines for the regeneration of connective tissue are shown in Table 1.

PLATELET-RICH PLASMA (PRP)

Platelets are small non-nucleated cells found in the peripheral blood that are involved in hemostasis¹¹⁷. Platelets are important in wound healing regulation through the release of a number of different cytokines, proteins and other biologically active molecules¹¹⁸. Platelet-rich plasma (PRP) is a blood product defined as a portion of the plasma fraction of autologous blood with an increased platelet concentration and an associated increase in growth factor concentration. PRP is obtained from autologous blood and prepared by simple centrifugation¹¹⁹. PRP has been shown to have a role in skin repair and healing¹²⁰ and it is becoming increasingly used in many regenerative medicine protocols^{121,122}.

The platelet alpha granules contain many growth factors including TGF- β , PDGF, insulin-like growth factors (IGF-1 and IGF-2), FGF, VEGF and epidermal growth factor (EGF)¹²³. The aforementioned growth factors have important regulatory effects on tissue homeostasis and MSC function which, as described earlier, have an important role in regenerative medicine and may even be an immunomodulatory route to treatment of coronavirus disease 2019 (COVID-19)¹²⁴. The immunomodulatory and anti-inflammatory properties of PRP are becoming increasingly important for the use of PRP in the treatment of musculoskeletal conditions and connective tissue diseases¹²⁵⁻¹²⁶.

THE USE OF GOLD-INDUCED CYTOKINES (GOLDIC[®]) IN THE REPAIR OF DAMAGED CONNECTIVE TISSUE

The development of GOLDIC[®] technology has enabled the production of autologous conditioned serum which is rich in anti-inflammatory cytokines (autologous gold-induced cytokines). GOLDIC[®]

is an *in vitro* gold treatment of autologous whole blood. The resultant gold-treated plasma is readministered to the patient without the presence of any residual gold compounds. Gold compounds have been used historically in the treatment of different inflammatory disorders, especially in musculoskeletal and rheumatic diseases, although these compounds are associated with many side effects¹²⁷⁻¹²⁹. *In vitro* studies have shown that incubation with gold particles inhibits catabolic factors, increases anti-catabolic and anabolic factors, and increases the level of gelsolin (GSN), which is a protein exerting an important role in cellular metabolism¹³⁰. The mechanism of action of GOLDIC[®] procedure has yet to be fully defined, but *in vitro* studies have shown a significant increase in plasma GSN levels in the autologous serum, as well as increased GSN levels in synovial fluid after intra-articular GOLDIC[®] injection therapy¹³¹. GSN is a cytoplasmic regulator of actin organization, which is responsible for the viscoelasticity of the cell cytoskeleton and regulates important cell functions including cell motility, phagocytosis, apoptosis¹³⁰. Plasma gelsolin (pGSN) can modulate pro-inflammatory pathways in rheumatoid arthritis, but local GSN levels in the affected joints are reduced even more than in plasma¹³². This considerably reduces the efficacy of endogenous GSN in rheumatoid arthritis and increases the importance of exogenous GSN (e.g., via GOLDIC[®] method) to treat inflammatory rheumatic diseases. It is clear that pGSN has a fundamental role in the modulation of pro-inflammatory responses, and, consistent with these functions, decreased pGSN levels have been detected in clinical conditions such as acute respiratory distress syndrome, sepsis, major trauma, prolonged neonatal hyperoxia, malaria, and liver injury^{133,134}. Moreover, the potential clinical utility of pGSN as a diagnostic tool has emerged in the aforementioned diseases, where circulating pGSN concentrations are below the normal values¹³⁴.

The first trial investigating the use of GOLDIC[®] method to treat lameness in horses showed a significant improvement in lameness-associated equine diseases following treatment¹³⁵. The first human clinical study using GOLDIC[®] method investigated the use of this technology in Achilles tendinopathy and found significant clinical and radiological (MRI) improvements¹³⁶. In another clinical study conducted in patients with osteoarthritis of the knee, intra-articular GOLDIC[®] injections

produced a rapid and sustained improvement in all symptoms, suggesting that GOLDIC® method represents a promising option for the conservative management of moderate to severe osteoarthritis of the knee¹³⁷. Most recently, an open-label, non-randomized, non-controlled study involving a heterogeneous patient population showed that GOLDIC® was safe and effective in the treatment of various systemic diseases such as allergies, fibromyalgia, psoriasis, rheumatoid arthritis, ulcerative colitis, polymyalgia rheumatica and osteoporosis¹³⁸.

CONCLUSIONS

In the regenerative technologies described in this review, a combination of connective tissue repair and regeneration processes occur following different interventions. The relative contributions of repair and regeneration are influenced by the proliferative capacity of the cells residing within a given tissue, by the ECM integrity and by the resolution or the chronic nature of injury and/or inflammation. Nevertheless, the most tissue-destructive diseases are caused by infections, autoimmune responses, trauma and other types of tissue injury, which may persist as chronic disease resulting in organ dysfunction and (often) organ failure. The persistence of disease leads to chronic inflammation, which is associated with the proliferation and activation of macrophages and lymphocytes, along with the production of a plethora of pro-inflammatory and pro-fibrotic growth factors and cytokines. Therefore, the intrinsic regulation of inflammation is critical for a proper tissue healing, repair and regeneration. In order to characterize the potency of the technologies described in this review, it is necessary to redefine such technologies according to the quality of the tissue regeneration that they produce:

- grade 1: complete restitution of damaged tissue without scar tissue or inflammation.
- grade 2: restitution of damaged tissue with low scar tissue content or inflammation.
- grade 3: restitution of damaged tissue with moderate scar tissue content or inflammation.
- grade 4: restitution of damaged tissue with high scar tissue content or inflammation.

In clinical settings, this grading can be defined by using histology and/or high-resolution MRI (3 Tesla MRI or higher). In order to avoid the ethical issues posed by invasive tissue biopsy, MRI image analysis seems to be the most appropriate procedure

for the assessment of repair and regeneration of damaged connective tissue. According to these criteria, GOLDIC® method may have the highest regenerative capacity and the lowest risk of side effects among all the regenerative procedures described in this review. In conclusion, it is likely that in future clinical practice a combination of technologies aimed to promote connective tissue repair and regeneration will offer the optimal treatment to patients suffering from various inflammatory diseases, which must always be our ultimate goal.

FUNDING:

No funding is declared for this article.

AUTHOR CONTRIBUTIONS:

Ulrich Schneider: Conception, design and drafting of the review, final approval for publication. William Murrell: Drafting of the review. Peter Hollands: Drafting and revision of the review, final approval for publication.

ORCID:

Ulrich Schneider: <https://orcid.org/0000-0002-2252-5188>

William Murrell: <https://orcid.org/0000-0001-8835-1806>

Peter Hollands: <https://orcid.org/0000-0003-4116-1954>

CONFLICT OF INTEREST:

Ulrich Schneider is the inventor of GOLDIC® and CEO of ArthroGen GmbH (Manufacturer of GOLDIC-Sets). William Murrell and Peter Hollands declare that they have no conflicts of interest to disclose.

REFERENCES

1. Stone RC, Pastar I, Ojeh N, Chen V, Liu S, Garzon KI, Tomic-Canic M. Epithelial-mesenchymal transition in tissue repair and fibrosis. *Cell Tissue Res* 2016; 365: 495-506. doi: 10.1007/s00441-016-2464-0.
2. Loi F, Córdova LA, Pajarinen J, Lin TH, Yao Z, Goodman SB. Inflammation, fracture and bone repair. *Bone* 2016; 86: 119-130. doi: 10.1016/j.bone.2016.02.020.
3. Nye HL, Cameron JA, Chernoff EA, Stocum DL. Regeneration of the urodele limb: a review. *Dev Dyn* 2003; 226: 280-294. doi: 10.1002/dvdy.10236. PMID: 12557206.
4. Chernoff EA, Stocum DL, Nye HL, Cameron JA. Urodele spinal cord regeneration and related processes. *Dev Dyn* 2003; 226: 295-307. doi: 10.1002/dvdy.10240.
5. Maden M. The evolution of regeneration - where does that leave mammals? *Int J Dev Biol* 2018; 62: 369-372. doi: 10.1387/ijdb.180031mm.
6. Seifert AW, Muneoka K. The blastema and epimorphic regeneration in mammals. *Dev Biol* 2018; 433: 190-199. doi: 10.1016/j.ydbio.2017.08.007.

7. Eming SA, Martin P, Tomic-Canic M. Wound repair and regeneration: mechanisms, signaling, and translation. *Sci Transl Med* 2014; 6: 265sr6. doi: 10.1126/scitranslmed.3009337.
8. Maeba T, Yonezawa T, Ono M, Tomono Y, Heljasvaara R, Pihlajaniemi T, Inagawa K, Oohashi T. Collagen XVIII deposition in the basement membrane zone beneath the newly forming epidermis during wound healing in mice. *Acta Med Okayama* 2019; 73: 135-146. doi: 10.18926/AMO/56649.
9. Ogawa R. Keloid and Hypertrophic Scars Are the Result of Chronic Inflammation in the Reticular Dermis. *Int J Mol Sci* 2017; 18: 606. doi: 10.3390/ijms18030606.
10. Klingberg F, Hinz B, White ES. The myofibroblast matrix: implications for tissue repair and fibrosis. *J Pathol* 2013; 229: 298-309. doi: 10.1002/path.4104.
11. Lu P, Takai K, Weaver VM, Werb Z. Extracellular matrix degradation and remodeling in development and disease. *Cold Spring Harb Perspect Biol* 2011; 3: a005058. doi: 10.1101/cshperspect.a005058.
12. Townsend SE, Gannon M. Extracellular Matrix-Associated Factors Play Critical Roles in Regulating Pancreatic β -Cell Proliferation and Survival. *Endocrinology* 2019; 160: 1885-1894. doi: 10.1210/en.2019-00206.
13. Wolf K, Te Lindert M, Krause M, Alexander S, Te Riet J, Willis AL, Hoffman RM, Figdor CG, Weiss SJ, Friedl P. Physical limits of cell migration: control by ECM space and nuclear deformation and tuning by proteolysis and traction force. *J Cell Biol* 2013; 201: 1069-1084. doi: 10.1083/jcb.201210152.
14. Muncie JM, Weaver VM. The Physical and Biochemical Properties of the Extracellular Matrix Regulate Cell Fate. *Curr Top Dev Biol* 2018; 130: 1-37. doi: 10.1016/bs.ctdb.2018.02.002.
15. Mettouchi A. The role of extracellular matrix in vascular branching morphogenesis. *Cell Adh Migr* 2012; 6: 528-534. doi: 10.4161/cam.22862.
16. Olczyk P, Mencner L, Komosinska-Vassev K. The role of the extracellular matrix components in cutaneous wound healing. *Biomed Res Int* 2014; 2014: 747584. doi: 10.1155/2014/747584.
17. Burgstaller G, Oehrle B, Gerckens M, White ES, Schiller HB, Eickelberg O. The instructive extracellular matrix of the lung: basic composition and alterations in chronic lung disease. *Eur Respir J* 2017; 50: 1601805. doi: 10.1183/13993003.01805-2016.
18. Frangogiannis NG. The extracellular matrix in myocardial injury, repair, and remodeling. *J Clin Invest* 2017; 127: 1600-1612. doi: 10.1172/JCI87491.
19. Paolillo M, Schinelli S. Extracellular Matrix Alterations in Metastatic Processes. *Int J Mol Sci* 2019; 20: 4947. doi: 10.3390/ijms20194947.
20. Edwards DN, Bix GJ. Roles of blood-brain barrier integrins and extracellular matrix in stroke. *Am J Physiol Cell Physiol* 2019; 316: C252-C263. doi: 10.1152/ajpcell.00151.2018.
21. McLeod CM, Mauck RL. High fidelity visualization of cell-to-cell variation and temporal dynamics in nascent extracellular matrix formation. *Sci Rep* 2016; 6: 38852. doi: 10.1038/srep38852.
22. Mouw JK, Ou G, Weaver VM. Extracellular matrix assembly: a multiscale deconstruction. *Nat Rev Mol Cell Biol* 2014; 15: 771-785. doi: 10.1038/nrm3902.
23. He L, Zhou J, Chen M, Lin CS, Kim SG, Zhou Y, Xiang L, Xie M, Bai H, Yao H, Shi C, Coelho PG, Bromage TG, Hu B, Tovar N, Witek L, Wu J, Chen K, Gu W, Zheng J, Sheu TJ, Zhong J, Wen J, Niu Y, Cheng B, Gong Q, Owens DM, Stanislauskas M, Pei J, Chotkowski G, Wang S, Yang G, Zegarelli DJ, Shi X, Finkel M, Zhang W, Li J, Cheng J, Tarnow DP, Zhou X, Wang Z, Jiang X, Romanov A, Rowe DW, Wang S, Ye L, Ling J, Mao J. Parenchymal and stromal tissue regeneration of tooth organ by pivotal signals reinstated in decellularized matrix. *Nat Mater* 2019; 18: 627-637. doi: 10.1038/s41563-019-0368-6.
24. Rippa AL, Kalabusheva EP, Vorotelyak EA. Regeneration of dermis: scarring and cells involved. *Cells* 2019; 8: 607. doi: 10.3390/cells8060607.
25. Maddaluno L, Urwyler C, Werner S. Fibroblast growth factors: key players in regeneration and tissue repair. *Development* 2017; 144: 4047-4060. doi: 10.1242/dev.152587.
26. Li X, He XT, Yin Y, Wu RX, Tian BM, Chen FM. Administration of signalling molecules dictates stem cell homing for in situ regeneration. *J Cell Mol Med* 2017; 21: 3162-3177. doi: 10.1111/jcmm.13286.
27. Strigini G, Ghidoni L, Quattrini F, Bellina G, Ciatti C, Fiazza C, Maniscalco P. Preliminary experience in the treatment of hip necrosis with BIOS screws associated with growth factors. *Acta Biomed* 2020; 91: 41-43. doi: 10.23750/abm.v91i1.7090.
28. Le ADK, Enweze L, DeBaun MR, Dragoo JL. Current Clinical Recommendations for Use of Platelet-Rich Plasma. *Curr Rev Musculoskelet Med* 2018; 11: 624-634. doi: 10.1007/s12178-018-9527-7.
29. Urits I, Capuco A, Sharma M, Kaye AD, Viswanath O, Cornett EM, Orhurhu V. Stem Cell Therapies for Treatment of Discogenic Low Back Pain: a Comprehensive Review. *Curr Pain Headache Rep* 2019; 23: 65. doi: 10.1007/s11916-019-0804-y.
30. Hu L, Yin C, Zhao F, Ali A, Ma J, Qian A. Mesenchymal Stem Cells: Cell Fate Decision to Osteoblast or Adipocyte and Application in Osteoporosis Treatment. *Int J Mol Sci* 2018; 19: 360. doi: 10.3390/ijms19020360.
31. Berthiaume F, Maguire TJ, Yarmush ML. Tissue engineering and regenerative medicine: history, progress, and challenges. *Annu Rev Chem Biomol Eng* 2019; 2: 403-430. doi: 10.1146/annurev-chembioeng-061010-114257.
32. Bogdanowicz DR, Lu HH. Designing the stem cell microenvironment for guided connective tissue regeneration. *Ann N Y Acad Sci* 2017; 1410: 3-25. doi: 10.1111/nyas.13553.
33. Zakrzewski W, Dobrzyński M, Szymonowicz M, Rybak Z. Stem cells: past, present, and future. *Stem Cell Res Ther* 2019; 10: 68. doi: 10.1186/s13287-019-1165-5.
34. Naji A, Eitoku M, Favier B, Deschaseaux F, Rouas-Freiss N, Suganuma N. Biological functions of mesenchymal stem cells and clinical implications. *Cell Mol Life Sci* 2019; 76: 3323-3348. doi: 10.1007/s00018-019-03125-1.

35. Vizoso FJ, Eiro N, Costa L, Esparza P, Landin M, Diaz-Rodríguez P, Schneider J, Perez-Fernandez R. Mesenchymal Stem Cells in Homeostasis and Systemic Diseases: Hypothesis, Evidences, and Therapeutic Opportunities. *Int J Mol Sci* 2019; 20: 3738. doi: 10.3390/ijms20153738.
36. Gneccchi M, Melo LG. Bone marrow-derived mesenchymal stem cells: isolation, expansion, characterization, viral transduction, and production of conditioned medium. *Methods Mol Biol* 2009; 482: 281-294. doi: 10.1007/978-1-59745-060-7_18.
37. Duchamp de Lageneste O, Julien A, Abou-Khalil R, Frangi G, Carvalho C, Cagnard N, Cordier C, Conway SJ, Colnot C. Periosteum contains skeletal stem cells with high bone regenerative potential controlled by Periostin. *Nat Commun* 2018; 9: 773. doi: 10.1038/s41467-018-03124-z.
38. Yang W, Guo D, Harris MA, Cui Y, Gluhak-Heinrich J, Wu J, Chen XD, Skinner C, Nyman JS, Edwards JR, Mundy GR, Lichtler A, Kream BE, Rowe DW, Kalajzic I, David V, Quarles DL, Villareal D, Scott G, Ray M, Liu S, Martin JF, Mishina Y, Harris SE. Bmp2 in osteoblasts of periosteum and trabecular bone links bone formation to vascularization and mesenchymal stem cells. *J Cell Sci* 2013; 126: 4085-4098. doi: 10.1242/jcs.118596.
39. Minteer D, Marra KG, Rubin JP. Adipose-derived mesenchymal stem cells: biology and potential applications. *Adv Biochem Eng Biotechnol* 2013; 129: 59-71. doi: 10.1007/10_2012_146.
40. Zupan J, Drobnič M, Stražar K. Synovium-Derived Mesenchymal Stem/Stromal Cells and their Promise for Cartilage Regeneration. *Adv Exp Med Biol* 2020; 1212: 87-106. doi: 10.1007/5584_2019_381.
41. Pantelic MN, Larkin LM. Stem Cells for Skeletal Muscle Tissue Engineering. *Tissue Eng Part B Rev* 2018; 24: 373-391. doi: 10.1089/ten.TEB.2017.0451.
42. Hollands P, Aboyeji D, Orcharton M. Dental pulp stem cells in regenerative medicine. *Br Dent J* 2018. doi: 10.1038/sj.bdj.2018.348.
43. Van SY, Noh YK, Kim SW, Oh YM, Kim IH, Park K. Human umbilical cord blood mesenchymal stem cells expansion via human fibroblast-derived matrix and their potentials toward regenerative application. *Cell Tissue Res* 2019; 376: 233-245. doi: 10.1007/s00441-018-2971-2.
44. Chandravanshi B, Bhonde RR. Human Umbilical Cord-Derived Stem Cells: Isolation, Characterization, Differentiation, and Application in Treating Diabetes. *Crit Rev Biomed Eng* 2018; 46: 399-412. doi: 10.1615/CritRevBiomedEng.2018027377.
45. Lanzoni G, Linetsky E, Correa D, Alvarez RA, Marttos A, Hirani K, Messinger Cayetano S, Castro JG, Paidas MJ, Efantis Potter J, Xu X, Glassberg M, Tan J, Patel AN, Goldstein G, Kenyon NS, Baidal D, Alejandro R, Vianna R, Ruiz P, Caplan AI, Ricordi C. Umbilical cord-derived mesenchymal stem cells for COVID-19 patients with acute respiratory distress syndrome (ARDS). *CellR4* 2020; 8: e2839. DOI: 10.32113/cellr4_20204_2839.
46. Lanzoni G, Linetsky E, Correa D, Messinger Cayetano S, Alvarez RA, Kouroupis D, Alvarez Gil A, Poggioli R, Ruiz P, Marttos AC, Hirani K, Bell CA, Kusack H, Rafkin L, Baidal D, Pastewski A, Gawri K, Leñero C, Mantero AMA, Metalonis SW, Wang X, Roque L, Masters B, Kenyon NS, Ginzburg E, Xu X, Tan J, Caplan AI, Glassberg MK, Alejandro R, Ricordi C. Umbilical cord mesenchymal stem cells for COVID-19 acute respiratory distress syndrome: A double-blind, phase 1/2a, randomized controlled trial. *Stem Cells Transl Med* 2021; doi: 10.1002/sctm.20-0472. Epub ahead of print
47. Macholdová K, Macháčková E, Prošková V, Hromadníková I, Klubal R. Latest findings on the placenta from the point of view of immunology, tolerance and mesenchymal stem cells. *Ceska Gynekol* 2019; 84: 154-160. English. PMID: 31238687.
48. Chen L, Qu J, Xiang C. The multi-functional roles of menstrual blood-derived stem cells in regenerative medicine. *Stem Cell Res Ther* 2019; 10: 1. doi: 10.1186/s13287-018-1105-9.
49. Patki S, Kadam S, Chandra V, Bhonde R. Human breast milk is a rich source of multipotent mesenchymal stem cells. *Hum Cell* 2010; 23: 35-40. doi: 10.1111/j.1749-0774.2010.00083.x.
50. Birmingham E, Niebur GL, McHugh PE, Shaw G, Barry FP, McNamara LM. Osteogenic differentiation of mesenchymal stem cells is regulated by osteocyte and osteoblast cells in a simplified bone niche. *Eur Cell Mater* 2012; 23: 13-27. doi: 10.22203/ecm.v023a02.
51. Munir H, Ward LSC, Sheriff L, Kemble S, Nayar S, Barone F, Nash GB, McGettrick HM. Adipogenic Differentiation of Mesenchymal Stem Cells Alters Their Immunomodulatory Properties in a Tissue-Specific Manner. *Stem Cells* 2017; 35: 1636-1646. doi: 10.1002/stem.2622.
52. Szychlińska MA, Stoddart MJ, D'Amora U, Ambrosio L, Alini M, Musumeci G. Mesenchymal Stem Cell-Based Cartilage Regeneration Approach and Cell Senescence: Can We Manipulate Cell Aging and Function? *Tissue Eng Part B Rev* 2017; 23: 529-539. doi: 10.1089/ten.TEB.2017.0083.
53. Richardson SM, Walker RV, Parker S, Rhodes NP, Hunt JA, Freemont AJ, Hoyland JA. Intervertebral disc cell-mediated mesenchymal stem cell differentiation. *Stem Cells* 2006; 24: 707-716. doi: 10.1634/stemcells.2005-0205.
54. Hevesi M, LaPrade M, Saris DBF, Krych AJ. Stem Cell Treatment for Ligament Repair and Reconstruction. *Curr Rev Musculoskelet Med* 2019; 12: 446-450. doi: 10.1007/s12178-019-09580-4.
55. Szaraz P, Gratch YS, Iqbal F, Librach CL. In Vitro Differentiation of Human Mesenchymal Stem Cells into Functional Cardiomyocyte-like Cells. *J Vis Exp* 2017; 9: 55757. doi: 10.3791/55757.
56. Rangel-Huerta E, Maldonado E. Transit-Amplifying Cells in the Fast Lane from Stem Cells towards Differentiation. *Stem Cells Int* 2017; 2017: 7602951. doi: 10.1155/2017/7602951.
57. Ferrin I, Beloqui I, Zabaleta L, Salcedo JM, Trigueros C, Martín AG. Isolation, Culture, and Expansion of Mesenchymal Stem Cells. *Methods Mol Biol* 2017; 1590: 177-190. doi: 10.1007/978-1-4939-6921-0_13.
58. Vega A, Martín-Ferrero MA, Del Canto F, Alberca M, García V, Munar A, Orozco L, Soler R, Fuertes JJ, Huguet M, Sánchez A, García-Sancho J. Treatment of Knee Osteoarthritis With Allogeneic Bone Marrow Mesenchymal Stem Cells: A Randomized Controlled Trial. *Transplantation* 2015; 99: 1681-1690. doi:10.1097/TP.0000000000000678.

59. Galipeau J, Sensébé L. Mesenchymal Stromal Cells: Clinical Challenges and Therapeutic Opportunities. *Cell Stem Cell* 2018; 22: 824-833. doi: 10.1016/j.stem.2018.05.004.
60. Radrizzani M, Soncin S, Lo Cicero V, Andriolo G, Bolis S, Turchetto L. Quality Control Assays for Clinical-Grade Human Mesenchymal Stromal Cells: Methods for ATMP Release. *Methods Mol Biol* 2016; 1416: 313-337. doi: 10.1007/978-1-4939-3584-0_19.
61. Brocher J, Janicki P, Voltz P, Seebach E, Neumann E, Mueller-Ladner U, Richter W. Inferior ectopic bone formation of mesenchymal stromal cells from adipose tissue compared to bone marrow: rescue by chondrogenic pre-induction. *Stem Cell Res* 2013; 11: 1393-406. doi: 10.1016/j.scr.2013.07.008.
62. Coleman SR, Lam S, Cohen SR, Bohluhi B, Nahai F. Fat Grafting: Challenges and Debates. *Atlas Oral Maxillofac Surg Clin North Am* 2018; 26: 81-84. doi: 10.1016/j.cxom.2017.10.006.
63. Tiryaki T, Condé-Green A, Cohen SR, Canikyan S, Kocak P. A 3-step Mechanical Digestion Method to Harvest Adipose-derived Stromal Vascular Fraction. *Plast Reconstr Surg Glob Open* 2020; 8: e2652. doi: 10.1097/GOX.0000000000002652.
64. Senesi L, De Francesco F, Farinelli L, Manzotti S, Gagliardi G, Papalia GF, Riccio M, Gigante A. Mechanical and Enzymatic Procedures to Isolate the Stromal Vascular Fraction From Adipose Tissue: Preliminary Results. *Front Cell Dev Biol* 2019; 7: 88. doi: 10.3389/fcell.2019.00088.
65. Tiryaki T, Canikyan S, Koçak P, Cohen S, Sterodimas A, Schlaudraff KU, Schefflan M, Hollands P. Adipose-derived Stromal Vascular Matrix (SVM): a new paradigm in regenerative medicine. *CellR4* 2021; 9: e3060.
66. Araña M, Mazo M, Aranda P, Pelacho B, Prosper F. Adipose tissue-derived mesenchymal stem cells: isolation, expansion, and characterization. *Methods Mol Biol* 2013; 1036: 47-61. doi: 10.1007/978-1-62703-511-8_4.
67. Gao F, Chiu SM, Motan DA, Zhang Z, Chen L, Ji HL, Tse HF, Fu QL, Lian Q. Mesenchymal stem cells and immunomodulation: current status and future prospects. *Cell Death Dis* 2016; 7: e2062. doi: 10.1038/cddis.2015.327.
68. Murphy MB, Moncivais K, Caplan AI. Mesenchymal stem cells: environmentally responsive therapeutics for regenerative medicine. *Exp Mol Med* 2013; 45: e54. doi: 10.1038/emm.2013.94.
69. Li F, Zhang K, Liu H, Yang T, Xiao DJ, Wang YS. The neuroprotective effect of mesenchymal stem cells is mediated through inhibition of apoptosis in hypoxic ischemic injury. *World J Pediatr* 2020; 16: 193-200. doi: 10.1007/s12519-019-00310-x.
70. Han B, Fan J, Liu L, Tian J, Gan C, Yang Z, Jiao H, Zhang T, Liu Z, Zhang H. Adipose-derived mesenchymal stem cells treatments for fibroblasts of fibrotic scar via down-regulating TGF- β 1 and Notch-1 expression enhanced by photobiomodulation therapy. *Lasers Med Sci* 2019; 34: 1-10. doi: 10.1007/s10103-018-2567-9.
71. Mazini L, Rochette L, Admou B, Amal S, Malka G. Hopes and Limits of Adipose-Derived Stem Cells (ADSCs) and Mesenchymal Stem Cells (MSCs) in Wound Healing. *Int J Mol Sci* 2020; 21: 1306. doi: 10.3390/ijms21041306.
72. Mannino G, Gennuso F, Giurdanella G, Conti F, Drago F, Salomone S, Furno DL, Bucolo C, Giuffrida R. Pericyte-like differentiation of human adipose-derived mesenchymal stem cells: an in vitro study. *World J Stem Cells* 2020; 12: 1152-1170. doi: 10.4252/wjsc.v12.i10.1152.
73. Caplan AI, Dennis JE. Mesenchymal stem cells as trophic mediators. *J Cell Biochem* 2006; 98: 1076-1084. doi: 10.1002/jcb.20886.
74. Byrne AM, Bouchier-Hayes DJ, Harmey JH. Angiogenic and cell survival functions of vascular endothelial growth factor (VEGF). *J Cell Mol Med* 2005; 9: 777-794. doi: 10.1111/j.1582-4934.2005.tb00379.x.
75. Wee P, Wang Z. Epidermal Growth Factor Receptor Cell Proliferation Signaling Pathways. *Cancers (Basel)* 2017; 9: 52. doi: 10.3390/cancers9050052.
76. Garnarczyk A, Jurzak M, Gojniczek K. Characteristic of the endogenous peptides--endothelins and their role in the connective tissue fibrosis. *Wiad Lek* 2008; 61: 126-134.
77. Engström W, Granerus M. Effects of fibroblast growth factors 19 and 20 on cell multiplication and locomotion in a human embryonal carcinoma cell line (Tera-2) in vitro. *Anticancer Res* 2006; 26: 3307-3310. PMID: 17094445.
78. Discher DE, Mooney DJ, Zandstra PW. Growth factors, matrices, and forces combine and control stem cells. *Science* 2009; 324: 1673-1677. doi: 10.1126/science.1171643.
79. Ferrara N, Gerber HP, LeCouter J. The biology of VEGF and its receptors. *Nat Med* 2003; 9: 669-676. doi: 10.1038/nm0603-669.
80. Schneider MD, Kirshenbaum LA, Brand T, MacLellan WR. Control of cardiac gene transcription by fibroblast growth factors. *Mol Reprod Dev* 1994; 39: 112-117. doi: 10.1002/mrd.1080390117.
81. Beier F, Leask TA, Haque S, Chow C, Taylor AC, Lee RJ, Pestell RG, Ballock RT, LuValle P. Cell cycle genes in chondrocyte proliferation and differentiation. *Matrix Biol* 1999; 18: 109-120. doi: 10.1016/s0945-053x(99)00009-8.
82. Fredriksson L, Li H, Eriksson U. The PDGF family: four gene products form five dimeric isoforms. *Cytokine Growth Factor Rev* 2004; 15: 197-204. doi: 10.1016/j.cytogfr.2004.03.007.
83. Anitua E, Andia I, Ardanza B, Nurden P, Nurden AT. Autologous platelets as a source of proteins for healing and tissue regeneration. *Thromb Haemost* 2004; 91: 4-15. doi: 10.1160/TH03-07-0440.
84. Shah P, Keppler L, Rutkowski J. A review of platelet derived growth factor playing pivotal role in bone regeneration. *J Oral Implantol* 2014; 40: 330-340. doi: 10.1563/AAID-JOI-D-11-00173.
85. Risau W, Flamme I. Vasculogenesis. *Annu Rev Cell Dev Biol* 1995; 11: 73-91. doi: 10.1146/annurev.cb.11.110195.000445.
86. Melincovici CS, Boşca AB, Şuşman S, Mărginean M, Mişu C, Istrate M, Moldovan IM, Roman AL, Mişu CM. Vascular endothelial growth factor (VEGF) - key factor in normal and pathological angiogenesis. *Rom J Morphol Embryol* 2018; 59: 455-467.
87. Vailhé B, Vittet D, Feige JJ. In vitro models of vasculogenesis and angiogenesis. *Lab Invest* 2001; 81: 439-452. doi: 10.1038/labinvest.3780252.

88. Aguilar-Cazares D, Chavez-Dominguez R, Carlos-Reyes A, Lopez-Camarillo C, Hernandez de la Cruz ON, Lopez-Gonzalez JS. Contribution of Angiogenesis to Inflammation and Cancer. *Front Oncol* 2019; 12: 1399. doi: 10.3389/fonc.2019.01399.
89. Barrientos S, Stojadinovic O, Golinko MS, Brem H, Tomic-Canic M. Growth factors and cytokines in wound healing. *Wound Repair Regen* 2008; 16: 585-601. doi: 10.1111/j.1524-475X.2008.00410.x.
90. Hu C, Jiang X. Role of NRP-1 in VEGF-VEGFR2-Independent Tumorigenesis. *Target Oncol* 2016; 11: 501-505. doi: 10.1007/s11523-016-0422-0.
91. Karaman S, Leppänen VM, Alitalo K. Vascular endothelial growth factor signalling in development and disease. *Development* 2018; 145: dev151019. doi: 10.1242/dev.151019.
92. Lacal PM, Graziani G. Therapeutic implication of vascular endothelial growth factor receptor-1 (VEGFR-1) targeting in cancer cells and tumor microenvironment by competitive and non-competitive inhibitors. *Pharmacol Res* 2018; 136: 97-107. doi: 10.1016/j.phrs.2018.08.023.
93. Kinashi H, Ito Y, Sun T, Katsuno T, Takei Y. Roles of the TGF- β -VEGF-C Pathway in Fibrosis-Related Lymphangiogenesis. *Int J Mol Sci* 2018; 19: 2487. doi: 10.3390/ijms19092487.
94. Turner N, Grose R. Fibroblast growth factor signalling: from development to cancer. *Nat Rev Cancer* 2010; 10: 116-129. doi: 10.1038/nrc2780.
95. Zakrzewska M, Marcinkowska E, Wiedlocha A. FGF-1: from biology through engineering to potential medical applications. *Crit Rev Clin Lab Sci* 2008; 45: 91-135. doi: 10.1080/10408360701713120.
96. Yu PJ, Ferrari G, Galloway AC, Mignatti P, Pintucci G. Basic fibroblast growth factor (FGF-2): the high molecular weight forms come of age. *J Cell Biochem* 2007; 100: 1100-1108. doi: 10.1002/jcb.21116.
97. Yamani A, Zdzalik-Bielecka D, Lipner J, Stańczak A, Piórkowska N, Stańczak PS, Olejkowska P, Hucz-Kalitowska J, Magdycz M, Dzwonek K, Dubiel K, Lamparska-Przybysz M, Popiel D, Pieczykolan J, Wiczorek M. Discovery and optimization of novel pyrazole-benzimidazole CPL304110, as a potent and selective inhibitor of fibroblast growth factor receptors FGFR (1-3). *Eur J Med Chem* 2021; 210: 112990. doi: 10.1016/j.ejmech.2020.112990.
98. Soulet L, Chevet E, Lemaitre G, Blanquaert F, Meddahi A, Barritault D. FGFs and their receptors, in vitro and in vivo studies: new FGF receptor in the brain, FGF-1 in muscle, and the use of functional analogues of low-affinity heparin-binding growth factor receptors in tissue repair. *Mol Reprod Dev* 1994; 39: 49-54. doi: 10.1002/mrd.1080390109.
99. Zinkle A, Mohammadi M. Structural Biology of the FGF7 Subfamily. *Front Genet* 2019; 10: 102. doi: 10.3389/fgene.2019.00102.
100. Barrientos S, Stojadinovic O, Golinko MS, Brem H, Tomic-Canic M. Growth factors and cytokines in wound healing. *Wound Repair Regen* 2008; 16: 585-601. doi: 10.1111/j.1524-475X.2008.00410.x.
101. Inampudi C, Akintoye E, Ando T, Briasoulis A. Angiogenesis in peripheral arterial disease. *Curr Opin Pharmacol* 2018; 39: 60-67. doi: 10.1016/j.coph.2018.02.011.
102. Allouche M. Basic fibroblast growth factor and hematopoiesis. *Leukemia* 1995; 9: 937-942.
103. Ornitz DM, Marie PJ. Fibroblast growth factor signaling in skeletal development and disease. *Genes Dev* 2015; 29: 1463-1486. doi: 10.1101/gad.266551.115.
104. Meng XM, Nikolic-Paterson DJ, Lan HY. TGF- β : the master regulator of fibrosis. *Nat Rev Nephrol* 2016; 12: 325-338. doi: 10.1038/nrneph.2016.48. PMID: 27108839.
105. Nickel J, Ten Dijke P, Mueller TD. TGF- β family co-receptor function and signaling. *Acta Biochim Biophys Sin (Shanghai)* 2018; 50: 12-36. doi: 10.1093/abbs/gmx126.
106. Suzuki-Inoue K, Tsukiji N. A role of platelets beyond hemostasis. *Rinsho Ketsueki* 2019; 60: 1283-1291. Japanese doi: 10.11406/rinketsu.60.1283.
107. Travis MA, Sheppard D. TGF- β activation and function in immunity. *Annu Rev Immunol* 2014; 32: 51-82. doi: 10.1146/annurev-immunol-032713-120257.
108. Shapouri-Moghaddam A, Mohammadian S, Vazini H, Taghadosi M, Esmaili SA, Mardani F, Seifi B, Mohammadi A, Afshari JT, Sahebkar A. Macrophage plasticity, polarization, and function in health and disease. *J Cell Physiol* 2018; 233: 6425-6440. doi: 10.1002/jcp.26429.
109. Ma J, Sanchez-Duffhues G, Goumans MJ, Ten Dijke P. TGF- β -Induced Endothelial to Mesenchymal Transition in Disease and Tissue Engineering. *Front Cell Dev Biol* 2020; 8: 260. doi: 10.3389/fcell.2020.00260.
110. Lodyga M, Hinz B. TGF- β 1 - A truly transforming growth factor in fibrosis and immunity. *Semin Cell Dev Biol* 2020; 101: 123-139. doi: 10.1016/j.semcdb.2019.12.010.
111. Sipos F, Galamb O. Epithelial-to-mesenchymal and mesenchymal-to-epithelial transitions in the colon. *World J Gastroenterol* 2012; 18: 601-608. doi: 10.3748/wjg.v18.i7.601.
112. Colak S, Ten Dijke P. Targeting TGF- β Signaling in Cancer. *Trends Cancer* 2017; 3: 56-71. doi: 10.1016/j.trecan.2016.11.008.
113. Dinarello CA. Historical insights into cytokines. *Eur J Immunol* 2007; 37: S34-45. doi: 10.1002/eji.200737772.
114. Clarke B. Normal bone anatomy and physiology. *Clin J Am Soc Nephrol* 2008; Suppl 3: S131-9. doi: 10.2215/CJN.04151206.
115. Adamopoulos IE. Inflammation in bone physiology and pathology. *Curr Opin Rheumatol* 2018; 30: 59-64. doi: 10.1097/BOR.0000000000000449.
116. Kwon H, Brown WE, Lee CA, Wang D, Paschos N, Hu JC, Athanasiou KA. Surgical and tissue engineering strategies for articular cartilage and meniscus repair. *Nat Rev Rheumatol* 2019; 15: 550-570. doi: 10.1038/s41584-019-0255-1.
117. Holinstat M. Normal platelet function. *Cancer Metastasis Rev* 2017; 36: 195-198. doi: 10.1007/s10555-017-9677-x.
118. Rodrigues M, Kosaric N, Bonham CA, Gurtner GC. Wound Healing. A Cellular Perspective. *Physiol Rev* 2019; 99: 665-706. doi: 10.1152/physrev.00067.2017.

119. Alves R, Grimalt R. A review of platelet-rich plasma: history, biology, mechanism of action, and classification. *Skin Appendage Disord* 2018; 4: 18-24. doi: 10.1159/000477353
120. Emer J. Platelet-Rich Plasma (PRP): Current Applications in Dermatology. *Skin Therapy Lett* 2019; 24: 1-6.
121. Everts P, Onishi K, Jayaram P, Lana JF, Mautner K. Platelet-rich plasma: new performance understandings and therapeutic considerations in 2020. *Int J Mol Sci* 2020; 21: 7794. doi: 10.3390/ijms21207794.
122. Fernández Viña M. , Camozzi L. , Spitaleri M. I. , Fernández Viña R. , Reinchisi G. Hyperconcentrated Platelet-Rich Plasma (High-PRP) for the treatment of a non-healing ulcer of the lateral malleolus: a case report and literature review. *CellR4* 2020; 8: e2873. DOI: 10.32113/cellr4_2020_2873.
123. Blair P, Flaumenhaft R. Platelet alpha-granules: basic biology and clinical correlates. *Blood Rev* 2009; 23: 177-89. doi: 10.1016/j.blre.2009.04.001.
124. Golchin A, Seyedjafari E, Ardeshirylajimi A. Mesenchymal Stem Cell Therapy for COVID-19: Present or Future. *Stem Cell Rev Rep* 2020; 16: 427-433. doi: 10.1007/s12015-020-09973-w.
125. Chen P, Huang L, Ma Y, Zhang X, Zhou J, Ruan A, Wang Q. Intra-articular platelet-rich plasma injection for knee osteoarthritis: a summary of meta-analyses. *J Orthop Surg Res* 2019; 14: 385. doi: 10.1186/s13018-019-1363-y.
126. Kaux JF, Emonds-Alt T. The use of platelet-rich plasma to treat chronic tendinopathies: a technical analysis. *Platelets* 2018; 29: 213-227. doi: 10.1080/09537104.2017.1336211.
127. Fuggle NR, Cooper C, Oreffo ROC, Price AJ, Kaux JF, Maheu E, Cutolo M, Honvo G, Conaghan PG, Berenbaum F, Branco J, Brandi ML, Cortet B, Veronese N, Kurth AA, Matijevic R, Roth R, Pelletier JP, Martel-Pelletier J, Vlaskovska M, Thomas T, Lems WF, Al-Daghri N, Bruyère O, Rizzoli R, Kanis JA, Reginster JY. Alternative and complementary therapies in osteoarthritis and cartilage repair. *Aging Clin Exp Res* 2020; 32: 547-560. doi: 10.1007/s40520-020-01515-1.
128. Situnayake RD, Grindulis KA, McConkey B. Long-term treatment of rheumatoid arthritis with sulphasalazine, gold, or penicillamine: a comparison using life-table methods. *Ann Rheum Dis* 1987; 46: 177-183. doi: 10.1136/ard.46.3.177.
129. Vuolteenaho K, Kujala P, Moilanen T, Moilanen E. Aurothiomalate and hydroxychloroquine inhibit nitric oxide production in chondrocytes and in human osteoarthritic cartilage. *Scand J Rheumatol* 2005; 34: 475-479. doi: 10.1080/03009740510026797.
130. Silacci P, Mazzolai L, Gauci C, Stergiopoulos N, Yin HL, Hayoz D. Gelsolin superfamily proteins: key regulators of cellular functions. *Cell Mol Life Sci* 2004; 61: 2614-2623. doi: 10.1007/s00018-004-4225-6.
131. Schneider U, Kumar A, Murrell W, Ezekwesili A, Yurdi NA, Maffulli N. Intra-articular gold induced cytokine (GOLDIC®) injection therapy in patients with osteoarthritis of knee joint: a clinical study. *Int Orthop* 2021; doi: 10.1007/s00264-020-04870-w.
132. Sadzyński A, Kurek K, Konończuk T, Zendzian-Piotrowska M. Gelsolin - variety of structure and functions. *Postepy Hig Med Dosw (Online)* 2010; 64: 303-309.
133. Suhler E, Lin W, Yin HL, Lee WM. Decreased plasma gelsolin concentrations in acute liver failure, myocardial infarction, septic shock, and myonecrosis. *Crit Care Med* 1997; 25: 594-598. doi: 10.1097/00003246-199704000-00007.
134. Bucki R, Levental I, Kulakowska A, Janmey PA. Plasma gelsolin: function, prognostic value, and potential therapeutic use. *Curr Protein Pept Sci* 2008; 9: 541-551. doi: 10.2174/138920308786733912.
135. Schneider U, Veith G. First Results on the Outcome of Gold-induced, Autologous-Conditioned Serum (GOLDIC) in the Treatment of Different Lameness-associated Equine Diseases 2013. *J Cell Sci Ther* 2013; 5: 151-157.
136. Schneider U, Wallich R, Felmet G, Murrell WD. Gold-Induced Autologous Cytokine Treatment in Achilles Tendinopathy. *ISAKOS 2017 G.L.* In: Canata (ed), *Muscle and Tendon Injuries 2017*; pp. 411-420.
137. Schneider U, Kumar A, Murrell W, Ezekwesili A, Yurdi NA, Maffulli N. Intra-articular gold induced cytokine (GOLDIC®) injection therapy in patients with osteoarthritis of knee joint: a clinical study. *Int Orthop* 2021; doi: 10.1007/s00264-020-04870-w. Epub ahead of Print.
138. Schneider U, Lotzof K, Murrell WD, Goetz von Wächter E, Hollands P. Safety and efficacy of systemically administered autologous Gold-Induced Cytokines (GOLDIC®) *CellR4* 2021; 9: e3132. doi: 10.32113/cellr4_20214_3132.